

Special Lectures ①

N

GLY1 and non-lysosomal degradation of *N*-glycans, free *N*-glycans and dolichol-linked oligosaccharides

Tadashi Suzuki

Glycometabolome Team,
Systems Glycobiology Research Group,
RIKEN Global Research Cluster



Dr. Tadashi Suzuki (Team Leader, Glycometabolome Team, RIKEN Global Research Cluster) received B.S. (1992), M.S. (1994) and D. Sc. (1997) from the Department of Biochemistry and Biophysics, Graduate School of Science, University of Tokyo, Japan. He then became a postdoctoral fellow at Department of Biochemistry and Cell Biology, State University of New York at Stony Brook (1997-2000). During this period he identified the gene encoding the cytoplasmic PNGase in yeast. In 2000 he became a Research Scientist/Research Assistant Professor at the same University. In December 2001 he returned to Japan to start an independent career as a Researcher, Precursory Research for Embryonic Science and Technology (PRESTO), Japan Science and Technology Agency (JST)(2001-2005). In February 2002 he became an Assistant Professor, University of Tokyo, and in January 2004 he moved to Osaka to serve as a Visiting Associate Professor, Osaka University. Since October 2007, he holds a current position as a Team Leader, Glycometabolome Team, Systems Glycobiology Research Group, RIKEN. He also serves as a Visiting Professor at Saitama University since 2010.

Dr. Suzuki is a recipient of Young Scientist Award, FCCA (Forum: Carbohydrate Coming of Age)(1996), Genzyme Award, Society for Glycobiology (1997), Young Investigator Award, the Japanese Biochemical Society (2005), Young Investigator Award, the Japanese Society of Carbohydrate Research (2008) and Glycobiology Significant Achievement Award, Society for Glycobiology (2016). He serves as an Executive Editor for *Biochimica Biophysica Acta-General Subjects* (2016-) and an editorial board member for *Glycobiology* (2016-).

The biosynthetic/processing pathways for *N*-glycans have been well characterized in mammalian cells. There are, however, issues that remain to be clarified concerning aspects of their degradation. While the molecular mechanism of the lysosomal degradation for *N*-glycoproteins has been well studied in relation to genetic disorders, evidence exists to suggest that there are also “non-lysosomal” degradation processes, which are now known to occur widely in eukaryotic cells.

The cytoplasmic peptide:*N*-glycanase (PNGase) is the enzyme involved in the non-lysosomal degradation of misfolded *N*-glycopro-

teins, a process called ERAD (ER-associated degradation)^{1),2)}. In 2012, a patient harboring mutations of PNGase gene (*NGLY1*) was first reported. Symptom of these patients includes developmental delay, multifocus epilepsy, involuntary movement and liver dysfunction. This report clearly demonstrated that the cytoplasmic PNGase plays a pivotal role in normal human development.

The past decade we intensively analyzed *Ngly1*-deficient mice and found that they are embryonic lethal in C57BL/6 (B6) background. Surprisingly, the additional deletion of *Engase*, encoding another cytosolic deglycosylating enzyme involved in the non-

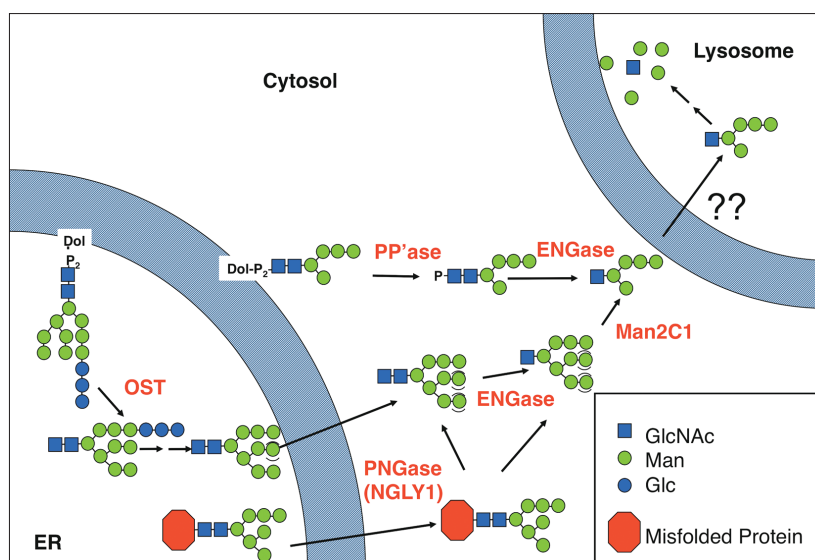


Figure 1 Non-lysosomal catabolism of *N*-glycoproteins, free *N*-glycans and dolichol-linked oligosaccharides in mammalian cells (Harada, et al., *Cell Mol Life Sci* 72, 2509-2533 (2015)). Dol: dolichol; ENGase: endo- β -*N*-acetylglucosaminidase; Man2C1: cytosolic α -mannosidase; OST: oligosaccharyltransferase; PNGase/NGLY1: peptide:*N*-glycanase

lysosomal degradation of *N*-glycoproteins called ENGase (endo- β -*N*-acetylglucosaminidase), resulted in the partial rescue of the lethality of the *Ngly1*-deficient mice³. Additionally, we also found that a change in the genetic background of B6 mice, produced by crossing the mice with an outbred mice strain (ICR) could rescue the embryonic lethality of *Ngly1*-deficient mice³. Viable *Ngly1*-deficient mice in a B6 and ICR mixed background, however, showed a very severe phenotype reminiscent of the symptoms of *NGLY1*-deficiency subjects. Again, many of those defects were strongly suppressed by the additional deletion of *Engase* in the B6 and ICR mixed background.

We also showed that in *Ngly1*-KO cells, ERAD process was compromised. Interestingly, not only delayed degradation but also the deglycosylation of a model substrate was observed in this cell. The unexpected deglycosylation was found to be mediated by ENGase. Surprisingly, the ERAD dysregulation in *Ngly1*-KO cells were restored by the additional KO of *Engase* gene. These observations collectively suggest that the ENGase represents one of the poten-

tial therapeutic targets for this genetic disorder⁴.

Besides degradation of *N*-glycoproteins, there are also novel pathways exist in cells for the catabolism of free *N*-glycans (FNGs); free oligosaccharides related to *N*-glycans, as well as dolichol-linked oligosaccharides (DLOs). The recent years we have also analyzed the following novel catabolic pathways for FNGs and DLOs; (i) oligosaccharyltransferase-mediated release of FNGs in the ER^{5,6}; (ii) pyrophosphatase acting on dolichol-linked oligosaccharides^{7,8}; and (iii) formation and catabolism of sialyl FNGs inside and outside of cells^{9,10}. Occurrence as well as biological importance of those novel pathways have been largely overlooked, partly because there are several genes for enzymes/transporters that remained to be identified. I have a firm belief that clarification of those novel pathway, which should constitute the one of the most basic biological processes, will lead to not only providing better insight into the process, but also understand the human health and disease. In this symposium, I will overview our recent findings on *NGLY1* and novel degradation pathways for FNGs/DLOs.

References

- 1) Suzuki T, Huang C, Fujihira H The cytoplasmic peptide:*N*-glycanase (NGLY1); structure, expression and cellular functions. *Gene* (577): 1-7, 2016
- 2) Suzuki T The cytoplasmic peptide:*N*-glycanase (PNGase) – basic science encounters a human genetic disorder. *J Biochem* (157): 23-34, 2015
- 3) Fujihira H, Masahara-Negishi Y, Tamura M, Huang C, Harada Y, Wakana S, Takakura D, Kawasaki N, Taniguchi N, Kondoh G, Yamashita T, Funakoshi Y, Suzuki T Lethality of *Ngly1*-knockout mice is partially rescued by the additional deletion of the *Engase* gene. *PLoS Genet* (13): e1006696, 2017
- 4) Huang C, Harada Y, Hosomi A, Masahara-Negishi Y, Seino J, Fujihira H, Funakoshi Y, Suzuki T, Dohmae T, Suzuki T Endo- β -*N*-acetylglucosaminidase forms *N*-GlcNAc protein aggregates during ER-associated degradation in *Ngly1*-defective cells. *Proc Natl Acad Sci USA* (112): 1398-1403, 2015
- 5) Harada Y, Buser R, Ngwa EM, Hirayama H, Aebi M, Suzuki T Eukaryotic oligosaccharyltransferase generates free oligosaccharides during *N*-glycosylation. *J Biol Chem* (288): 32673-32684, 2013
- 6) Harada Y, Masahara-Negishi Y, Suzuki T Cytosolic-free oligosaccharides are predominantly generated by the degradation of dolichol-linked oligosaccharides in mammalian cells. *Glycobiology* (25): 1196-1205, 2015
- 7) Harada Y, Nakajima K, Masahara-Negishi Y, Freeze HH, Angata T, Taniguchi N, Suzuki T Metabolically programmed quality control system for dolichol-linked oligosaccharides. *Proc Natl Acad Sci USA* (110): 19366-19371, 2013
- 8) Harada Y, Huang C, Yamaki S, Dohmae N, Suzuki T Non-lysosomal degradation of phosphorylated oligosaccharides is initiated by the cytosolic endo- β -*N*-acetylglucosaminidase. *J Biol Chem* (291): 8048-8058, 2016
- 9) Seino J, Wang L, Harada Y, Huang C, Ishii K, Mizushima N, Suzuki T Basal autophagy is required for the efficient catabolism of sialyl oligosaccharides. *J Biol Chem* (288): 26898-26907, 2013
- 10) Seino J, Fujihira H, Nakakita S, Masahara-Negishi Y, Miyoshi E, Hirabayashi J, Suzuki T Occurrence of sialyl oligosaccharides related to *N*-glycans (sialyl FNGs) in animal sera. *Glycobiology* (26): 1072-1085, 2016